

# Frequency of micronuclei and apoptosis in exfoliated buccal cells in chronic in flammation of oral mucosa

Fadil A. AL-QuraisheReceived 3-2-2016,Accepted 16-2-2016,Published 13-10-2016

#### Abstract

Chronic inflammations of the oral mucosa (CIOM) induced by biological, chemical, and physical factors and are in turn associated with an increased risk of oral cancers. The objective of the current study was the investigate the spontaneous genetic damageusingthe micronuclei (MN) and apoptosis test on exfoliated cells from (CIOM) patients. Cytological tests included, in addition to (MN), counting apoptosis (karyorrhexis, condensed chromatin, and, pyknosis). The study sample comprised 42 individuals aged (20-40) years including twenty two patients (13 males and 9 females) revealed (CIOM) and the another 20 (12 males and 8 females) were healthy control group. MN frequency was significantly increased (P<0.001) in buccal mucosa cells obtained from lesion area (LA) and normal area (NA) from chronic inflammation patients with mean value  $(7.53 \pm 1.49)$  and  $(5.02 \pm 1.49)$ 1.47) respectively, when compared with the healthy control group ( $2.40 \pm 1.40$ ). Lower frequency of apoptosis observed in (LA) from chronic inflammation patients compared with control group, was (P=0.06) more over apoptosis was significant increased (P<0.001) in NA of chronic inflammation patients when compared with cells of control group. The MN was regarded with the gender, age, mouthwashes uses, and, oral hygiene of the patients. The frequency of MN was significantly higher in the females than males in both patients and control groups (P < 0.01), whereas non-significant differences in MN occurrence was observed in relation to age.

However MN frequency was significantly higher in poor oral hygiene and mouthwash users (P<0.01) patients group but shown non-significant differences in poor and good oral hygiene in control group. Also the significantly higher frequency was increased MN in mouthwashes user in control group, when compared with non-users. The results suggest that increased MN and decreased apoptosis in the long periods chronic inflammation of the oral mucosa patients could be consider as a useful diagnostic bio-monitoring assay to prevent the risk from transformation of chronic inflammation to pre cancers lesions.

Key Words :Micronuclei ,Apoptosis, Chronic inflammation, Oral mucosa exfoliated.

## Introduction

Buccal cells form the first barrier of the inhalation or ingestion route and are capable of metabolizing and inflammation proximate carcinogens to reactive products (1-2-3). There-for, it could be argued the oral epithelial cells represent a preferred target sit for early genotoxic events induced bv carcinogenic agents entering the body via inhalation and ingestion (4). The oral epithelium maintains itself by continuous cell renew, whereby new cells produced in the basal layer by mitosis migrate to the surface replacing those that are shed (5). How-ever the oral mucosa, in addition to its unique micro environmental niche fueled by food residues; microbial flora; and saliva, has also been recognized for its sensitivity to inflammation, fibrosis, response proneness to drug, and carcinogenic and mutagenic agents (6-7). Oral mucosa mainly exposed to sources inflammation, the of wide spread and inflammation are include microbial and viral infections, exposure to allergens, toxic chemicals, consumption of alcohol and tobacco (8-9).

In general, the longer the inflammation persists, the higher risk of cancer. Two stages of inflammation exist; acute and chronic inflammation. Acute inflammation is an initial stage of inflammation, which is mediated through the activation of innate immune system, this type of inflammation persists only for a short time. the second stage of inflammation, or chronic inflammation, sets in and may predispose the host to various chronic illnesses, including

cancer (10). During inflammation, mast cells and leukocytes are recruited to the site of damage, which lead to a "respiratory burst" due to an increased uptake of oxygen, and thus. an increased release and accumulation of reactive oxygen species ROS at the site of damage (11-12) when (ROS) produced over long time, and thus significant damage may occur to the cell structure and functions and may induced somatic mutations and neoplastic transformation in site of chronic inflammation(13-14).

epidemiology Molecular research focuses on the biomarkers of exposure cytogenetic (e.g., endpoints- chromosomal aberrations, micronuclei. sister chromatid exchanges, and, apoptosis) (5-15-16). MN assay has been applied to evaluate chromosomal damage for biological population monitoring of human exposed to variety of mutagenic and carcinogenic agents (17-18-19).

Buccal cells not only offer the clinician opportunities for early diagnosis but also provide a unique model for mutation research that permits correlating genetic alteration with histopathologic changes and for drug discovery investigations (4-20). The objective of the current study was to investigate the spontaneous genetic damage in exfoliated cells of the buccal mucosa from chronic inflammation oral mucosa patients and healthy controls by the abnormalities cells (MN and apoptosis ) assay with exfoliated buccal mucosa cells regarding the factors that might affect MN and apoptosis frequency (i.e., ch-ronic inflammation,

gender, age, oral hygiene and

### Materials and Methods

Samples: Twenty- two patients (13 males and 8 females) revealed clinical signs of chronic inflammation of the oral mucosa were used in this study.In addition,20 healthy (12 males and 8 females) were used and as control group. considered The patients and control group individuals were aged (20- 40) year's. A written consent was taken from each individual and the samples were taken from AL-Specialization Muthanna Dental Center. The period of study was extended for 6 months .This study was hospital approved by the ethical committee approved the human study.

Micronuclei and Apoptosis Assay: The samples of exfoliated cells were collected from buccal mucosa in area with lesions and area without lesions (normal area )from patients and from control group. The participants were also asked to rinse the oral cavity for 1 minute with 10 ml. of sterilized distilled water and exfoliated cellsof buccal mucosa were obtained by a light and gently pressure was applied, while scraping the buccal mucosa with cytobrush moistened with buffer (21-22-23) for each individual, cytobrush used to collected buccal cells was shaken in a centrifuge tube containing saline solution( Hank's basic or other buffer solution) to release the cells and the tube was then centrifuged to wash

### **Results :**

The sample characteristics ate represented, as that shown in The (table-1), the mean age  $\pm$  SD of the

mouthwash use).

the cells in the buffer solution twice, was discarded and The supernatant pellets were re-suspended in 0.75 M Kcl and fixed by a cold methanol- glacial acid mix (3:1), then the tube stored at room temperature until investigation of MN and apoptosis. Slides were prepared by adding one drop of fixed cells solution onto center of clean glass slides and spreading on the slides. Afterwards ,the glass slides wasdried in air. Staining was carried out with 2% giemsa solution for a period of 10 minutes. Afterwards, the glass slide was rinsed with distilled water and dried in the air. The criteria of MN evaluation were those suggested by This study was approved by the hospital ethical committee Tolbert et.al.(1992) and Titenko-Holland 1998 (12).abnormalities, Screened for cell addition to counting MN, nuclear alterations suggestive of apoptosis were also investigated under oil immersion followed by (1000x), lens phase contrast microscopy for counting of MN and apoptosis according to established methods (22- 23- 24). At least 1000 epithelial cells per individual intact were scored to achieve the average percent of MN cells and apoptosis. Statistical analysis was done by SPSS version 15, comparison by (ANOVA-LSD) and correlation by Spearman correlation.

whole sample was  $(35.22\pm 3.07)$  for (CIOM) patients and healthy control with means were  $(38.55\pm 3.49)$  and  $(34.64\pm 2.44)$  respectively which

| characteristic | Gro             | P value  |         |
|----------------|-----------------|----------|---------|
|                | Patients        | control  |         |
|                | <b>No.</b> = 22 | No. = 20 |         |
| Gender         | No.%            | No.%     |         |
| Female         | 9 40.9%         | 8 40%    | P<0.01* |
| Male           | 13 59.1%        | 12 60%   | 0.040   |
| ·              | Oral hy         | giene    |         |
| Good           | 3 13.7%         | 5 25%    |         |
| Poor           | 19 86.3%        | 15 75%   |         |
|                | Mouthwa         | sh use   |         |
| Use            | 5 22.7%         | 9 45%    |         |
| Non            | 17 77.3%        | 11 55%   | 0.012   |
|                | Age (ye         | ars)     |         |
| 20-30          | 14 63.6%        | 9 45%    |         |
| 30-40          | 8 36.4%         | 11 55%   | 0.013   |

indicated no statistically significant

differences between the groups.

\* Significant p<0.01

Micronuclei analysis , results showed (Table -2, Fig-2a,2b) the MN frequency was significantly higher (P<0.001) in buccal mucosa cells obtained from lesion area (LA) than those of normal area(NA) in chronic inflammation

patients with mean SD value (7.53  $\pm$ 1.49) and (5.02  $\pm$  1.47) respectively in comparison with their healthy control group with mean  $\pm$  SD was (2.40 $\pm$ 1.40).

| Table 2: Micronuclei cell frequency in buccal mucosa exfoliated cells of (CIOM) |
|---|
| patients and healthy controls.  |

| Subject         | No.of      | MN-    | MN(%) (Mean      | Comparison   | P value   |
|-----------------|------------|--------|------------------|--------------|-----------|
|                 | individual | Range  | SD/1000cells)    |              |           |
| Patients Lesion | 22         | 1 - 20 | 7.53 ±0.78       | LA versus    | P <0.001* |
| area (LA)       |            |        |                  | control      |           |
| Patients Normal | 22         | 1 - 18 | $5.02 \pm 0.533$ | LA versus NA | P =0.796  |
| area (NA)       |            |        |                  |              |           |
| Control         | 20         | 0 - 6  | 2.40 ±0.40       | NA versus    | P <0.001* |
|                 |            |        |                  | control      |           |

\*Significant at P<0.001 compared with control

LA with control Significant.NA with control Significant.LA with NA Non-Significant.

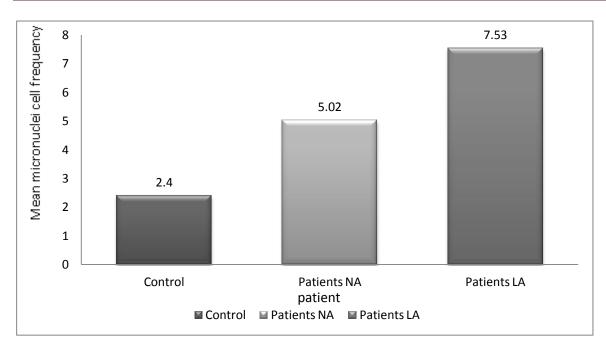


Figure 2a:- shows Micronuclei cell frequency in buccal mucosa exfoliated cells of chronic inflammation oral mucosa patients and healthy controls.

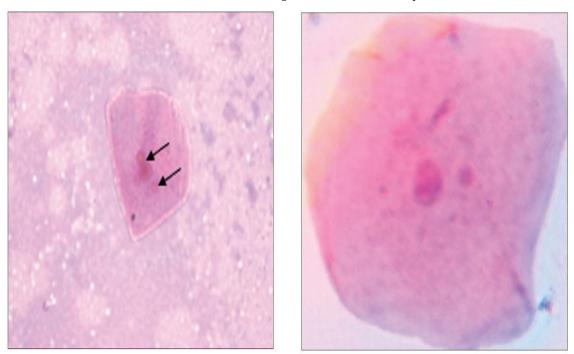


Figure 2 b: shows epithelium of buccal cell

Apoptosis, degenerative nuclear alterations indicative apoptosis of includes (karyorrhcetic, observed. condensed chromatin, and, pyknosis), As observed in (Table 3), the frequency of apoptosis did not show significant difference cells between buccal obtained of chronic from (LA)

inflammation patients compared with cells of healthy control group (P=0.061), whereas apoptosis frequency was significantly increased in buccal cells obtained from (NA) of chronic inflammation patients compared with cells of control group (P<0.001).

| Subject         | No. of     | Total cells | Karyorrhcet | Condensed | Pyknosis | P value    |
|-----------------|------------|-------------|-------------|-----------|----------|------------|
|                 | individual |             | ic          | chromatin |          |            |
| Patients Lesion | 22         | 48.01       | 322         | 439       | 170      | 0.061      |
| area (LA)       |            |             |             |           |          |            |
| Patients        | 22         | 51.92       | 393         | 580       | 136      | <b>P</b> < |
| Normal area     |            |             |             |           |          | 0.001*     |
| (NA)            |            |             |             |           |          |            |
| Control         | 22         | 44.32       | 402         | 429       | 77       | 0.063      |

 Table 3 : Frequency apoptosis in exfoliated buccal cells of chronic inflammation oral mucosa patients and healthy controls.

\*Significant at p< 0.001 compared with control.

The comparison mean  $\pm$ SD apoptosis analysis in buccal mucosa cells shown in (Table 4 and fig 4), there was no significant difference in the frequency of apoptosis between cells obtained from (LA) in patients compared with apoptosis in cells of the with control group mean  $\pm$ SD  $(27.81 \pm 3.45)$  $(28.08 \pm 4.65)$ and

respectively It was less apoptosis mean in the (LA) cells from patients than cells of control group. But, it showed significant increase in the apoptosis (P<0.001) in cells obtain from (NA) in patients compared with cells of the control group with mean  $(32.49\pm5.60)$ and  $(28.08\pm4.65)$  respectively.

Table 4: Comparison Apoptosis analysis(Karyorrhcetic, condensed chromatin and pyknosis) in buccal mucosa exfoliated cells of chronic inflammation oral mucosa patient sand healthy controls

| controis.   |           |           |                     |                    |             |  |
|-------------|-----------|-----------|---------------------|--------------------|-------------|--|
| Subject     | No. of    | Apoptosis | Apoptosis (%) (Mean | <b>Total cells</b> | P value     |  |
|             | individul | number    | ±SD / 100 cells)    |                    |             |  |
| Patients    | 22        | 931       | 27.81 ±3.45         | 48.01              | 0.478       |  |
| Lesion area |           |           |                     |                    |             |  |
| Patient     | 22        | 1.109     | $32.49 \pm 6.60$    | 51.92              | P < 0.001** |  |
| Normal area |           |           |                     |                    |             |  |
| control     | 20        | 908       | $28.08 \pm 4.65$    | 44.32              | 0.579       |  |
|             |           |           |                     |                    |             |  |

\*\* Significant at p < 0.001 compared with control.

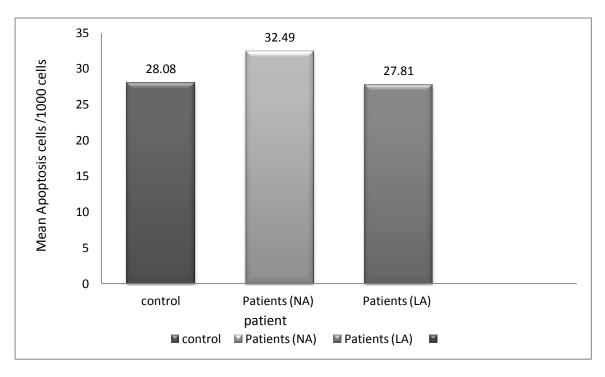


Figure 4:shows the Comparison between the apoptosis in buccal mucosa exfoliated cells of chronic inflammation oral mucosa patients and healthy controls.

The comparison between patients cell control group, revealed, nonsignificant differences in MN mean value occurrence, in relation to age. However MN occurrence was significantly higher in females, poor oral hygiene and mouthwash users (P < 0.01). Table (5) and figure (5),showed the significantly increased MN mean  $\pm$  SD in females than males in both buccal cells from patients and control groups with mean ± SD, in females  $(2.99\pm1.48)$  and  $(8.61\pm3.22)$ , when in males  $(1.88 \pm 1.33)$ and  $(6.45\pm2.99)$  respectively. However, MN occurrence was significantly higher in poor oral hygiene and mouth- wash users (P<0.01) in chronic inflammation patients, whereas the frequency of MN was significantly higher in mouthwash use of control group, when compared with non-user with mean  $\pm$ SD  $(2.78 \pm 1.46)$  $(1.99 \pm 1.88)$ and respectively.

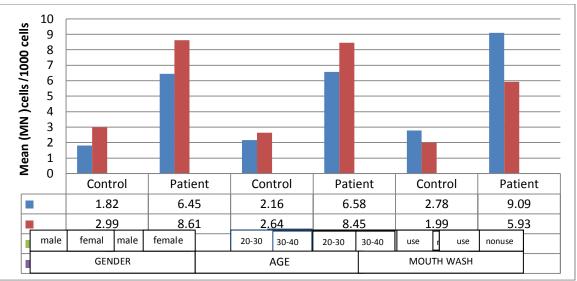


Figure 5: shows The comparison Mean (MN%) cells inbuccal exfoliated cells of chronic inflammation oral mucosa patients and healthy controls, by gender, age, and mouth wash use a variables.

Table 5: shows The comparison between the value Mean (MN%) in buccal mucosa exfoliated cells of chronic inflammation oral mucosa patients and healthy controls, by gender, and mouthwash use a variables.

|                |        | MN (Mean S      |             |           |
|----------------|--------|-----------------|-------------|-----------|
| Characteristic |        | Control         | Patients    | P value   |
|                | Male   | $1.82 \pm 1.33$ | 6.45± 2.99  |           |
| Gender         | Female | 2.99± 1.48*     | 8.61± 3.22* | P < 0.01* |
|                | Good   | 2.11 ± 1.97*    | 6.58± 2.69  |           |
| Oral hygiene   | Poor   | 2.64 ±1.24*     | 8.45± 3.48* | P < 0.01* |
|                | Use    | 2.78 ±1.46*     | 9.09± 4.33* |           |
| Mouthwash use  | Non    | 1.99± 1.88      | 5.93± 2.75  | P < 0.01* |

Significantly different compared between characteristics in groups P< 0.01.\*

#### Discussion

The micronuclei, is a recently upgraded topic, especially in the oral cancer filed (25), the cytogenetic biomarker of genome damage (e. g., MN, nuclear bud) and cell death (e. g. ,karyolysis, apoptosis) can be observed in both the lymphocytes and buccal mucosa cellssystems, and provide a more comprehensive assessment of genome damage of these MN and apoptosis in the context of cytotoxicity and cytostatic effects (26-27). The significantly higher frequency of MN in exfoliated oral buccal cells from chronic inflammation patients with oral mucosa observed in this study, corroborate the results from some studies, The hypothesis of a direct association between the frequency of MN in target or surrogate tissues and cancer development, is supported by the finding like in clear increase in the frequency of MN in target tissue as well in peripheral lymphocytes in cancer patients (6-25-28-29). Al though this

association has been described by other worker indicated the MN in exfoliated oral epithelial cells represent a preferred target site for early genotoxic events induced by carcinogenic agents (30-27) studies have shown Various the correlation of frequency of MN an severity of this genotoxic damage (29-27).Also the results are in agreement with those of other workers, the site chronic suggest in of inflammation such as when occur in the oral mucosa .Indeed cancer initiation and progression has been linked to oxidative stress by increasing mutation or inducing DNA DNA damage ,genome instability and cell proliferation (27-31-32-33).

The increase of MN frequency had been found associated with gender, we noted an increase in MN in female of chronic inflammation of oral mucosa patients, and in control groups, this increase may occur according to normally female have inactivated Xchromosome (Barr body) occupy site neighbor to nucleus in cytoplasmic of the cells. that lead to increase the counting number of MN in female samples than male. these result covenant with other workers indicated increased frequency of MN in female than male (19-34-35).

The significant increase of MN in mouthwash users in patients and control groups was also observed in this study that evaluated the genotoxic effect of risk factor for oral cancer development these occur according to components of mouthwash, ethanol is still a component of oral- care products (36–37). However, recent study showed that the genotoxicity of mouthwashes is caused by ethanol and not by any other ingredient (38), the mucosa may be damaged by ethanol, which leads to the stimulation of cell regeneration and genetic changes may then cause the development dysplasia, of or leukoplakia, finally and, cancer (38). The possibility of damage to the oral muosa exists with the use of mouthwashes (39). the risk was confined to users of mouthwash high in alcohol content (>25% Vol.) (40-41), It may from these results. be hypothesized that the use of mouthwashes could have a threshold for adverse effects.

The poor oral hygiene increased frequency, these observations MN suggest that there may be an influence on risk for oral cancer (39), poor oral hygiene increased the acetaldehyde production in oral micro flora, however, the metabolic acetaldehyde production directly affect the mucosa by alcohol dehydrogenase salivary and the acetaldehyde represents mostly microbial acetaldehyde formation in the oral cavity which increases the risk of genotoxicity to oral mucosa and may lead to genotoxic events induced by carcinogenic agents (25-42).

Therefore the lower frequency of apoptosis observed in lesion of patients group, indicate that with evolution of malignant transformation. the apoptotic response fails, as also bserved in precursor lesions of cervical (43–44). possible cancer The mechanisms include induction of genomic instability, alteration in epigenetic events and subsequent in appropriate gene expression, enhanced proliferation of initiated cells, resistance

to apoptosis, aggressive tumor new vascularization , invasion through tumor–associated basement membrane and metastasis (29–44–45).

The results, obtained in the present study showed that oral chronic inflammation is associated with higher frequency of chromosomal damage suggests and that apoptosis in decreased frequency is associated with evolution of malignant transformations.

## **Reference:**

1-Autrup H, Seremet T, Arenholt D, Dragsted L, Jepsen A. Metabolism of benzo[a] pyrene by cultured ratand human buccal mucosa cells. Carcinogenesis 1985; 6:1761–5.

2- Liu y, Sundqvist K, Belinsky SA, Castonguay A, Tjalve H, Grafstrom RC. Metabolism and macromolecular interaction at the tobacco-specific carcinogen 4-(methylnitrosamino)– 1-(3-pyridyl)– 1- butanone in cultured explants and epithelial cells of human buccal mucosa. Carcinogenesis 1993; 14: 2383-8.

3- VondracekM, Xi Z, LarssonP, BakerV, Mace K, Pfeifer A,et. al. Cytochrome P450 expression and related metabolism in human buccal mucosa. Carcinogenesis 2001;22:481-8. 4- Proia NK , Paszkiewicz GM, Nasca MA, Nersesyan A , Kundi M, Alefie K, et.al. Smoking and smokeless tobaccoassociated human buccal cancer- A review . Cancer Epidemiol Biomarkers Prev 2006; 15:1061-71.

5- Tolbert PE, Shy CM,Allen JW. Micronuclei and other nuclear anomalies in buccal smear : Methods development . Mutat Res 1992 ; 271: 69-77. Additionally, they suggest that mouthwashes and poor oral hygiene are effective in inducing chromosomal damage.

In conclusion, the degenerative nuclear alteration could be consider as indicative of apoptosis .Beside MN can be se as a useful assay for biomonitoring to increased risk of transformation of chronic inflammation of oral mucosa to cancer.

6- Holland N, Bolognesi C, Kirsch-Volders M, Bonassis S, Zeiger E, Knasmueller S, et. al. The micronuclei assay in human buccal cells as a tool for biomonitoring DNA damage . Mutat Res 2008 ; 659 : 93-108.

7- Boffetta P. Molecular epidemiology . J Inter Med 200; 248: 447- 54 .

8- Nair U, Obe G, Nair J, Maru GB, Bhide SV, Piepa R, et. al. Evaluation of frequency of micronucleated oral mucosa cells as a marker for genotoxic damage in chewers of betel quid with or without tobacco. Mutat Res 1999; 261: 163-8.

9- Palve DH, Tupkari JV .Clinicopathological correlation of micronuclei in oral squamous cells carcinoma by exfoliative cytology. J Oral MaxillofacPathol 2008; 1: 2-7.

10- Tian D , Ma H, Feng Z, Xia Y, Le XC, Ni Z, et. al. Analysis of micronuclei in exfoliated epi-thelial cells from individual chronically exposed to arsenic via drinking water in inner Mongo-lia, China. J Toxicol Environ Health A 2001 ; 64 : 473- 84.

11- Stich HF,Stich W, Rosin MP, Vallejera MO. Use of micronucleus test to monitor the effect of vitamin A, beta-carotene and canthaxanthin on the buccal mucosa of betel nut/ tobacco chewers. Int J Cancer 1984; 34: 745-50. 12- Titenko-Holland N, Jacob RA, Shang N, Balaraman A, Smith MT. Micronuclei in lymph- oocytes exfoliated buccal cells of postmenopausal women with dietary changes in folate . Mutat Res 1998; 417: 101-14.

13- Sarto F, Finotto S, Giacomelli L, Mazzotti D, Tomanin R, Levis AG. The micronucleus assayin exfoliated cells of human buccal mucosa. Mutagenesis 1987; 2: 11-7.

14- Tolbert PE, Shy CM, Allen JW. Micronuclei and other nuclear anomalies in buccal smears: A filed test in snuff users. Am J Epidemiol 1991; 134: 840-50.

15- Stich Hf, San RH, Rosin MP. Adaptation of the DNA– repair and micronucleus test to human cell suspensions and exfoliated cells. Ann NY AcadSci 1983; 407: 93-105.

16- Fenech M, Crott JW. Micronuclei, nucleoplasmic bridges and nuclear buds induced in folic acid deficient human lymphocytes–evidence for breakage fusion - bridge cycles in the cytokinesis–block micronuclei assay. Mutat Res 2002; 504 : 131-6.

17- Martinez FD. Gene –environment interactions in asthma and allergies: A newparadigm to understand disease causation. Immunol Allergy Clin North Am 2005; 25: 709 -21.

18- Pastors S, Creus A, Xamena N, Siffe IC, Marcos R. Occupational exposure to pesticides and cytogenetic damage: Results of a Hungarian population study using the micronucleus assay in lymphocytes and buccal cells. Environ Mol Mutagen 2002;40: 101-9.

19- Pastor S, Gutierrez S, Creus A, Xamena N, Piperakis S, Marcos R. Cytogenetic analysis of Greek farmers using the micronucleus assay in peripheral lymphocytes and buccal cells. Mutagenesis 2001; 16: 539 -45.

20- Holland N, Harmatz P, Golden D, Hubbard A, Wu YY, Bae J, et. al. Cytogenetic damage in blood lymphocytes and exfoliated epithelial cells of children with inflammatory bowel dis-ease . Pediatr Res 2007; 61: 209–14.

21- Chen C, Arjomandi M, Qin H, Balmes J, Tager I, Holland N. Cytogenetic damage in buccal epithelia and peripheral lymphocytes of young healthy individuals exposed to ozon. Mutagenesis 2006; 2: 131-7.

22- Speit G, Schmid O. Local genotoxic effects of formaldehyde in human measured by the micronucleus test with exfoliated epithelial cells. Mutat Res 2006; 613: 1–9.

23- Casartelli G, Bonatti S, De Ferrari DM, Scala M, Merea P, Margarino G, Micronu-clei frequency al. in et. cells in normal exfoliated buccal precancerous lesions and mucosa, squam-ous cell carcinoma. Anal Quant CytolHistol 2000; 22: 486-92.

24- Bonassi S, Neri M, Puntoni R. Validation of biomarkers as early predictors of disease. Mutat Res 2001; 480:349–81.

25- Jadhav K, Gupta N and Ahmed M BR. Micronuclei: An essential biomarker in oral exfoli-ated cells for grading of oral squamous cell carcinoma. J Cytol . 2001; 28(1):7–12. 26- Souza JP, Cerquerira E de M, Meireles JR. Chromosome damage, apoptosis, and necrosis in exfoliated cells of oral mucosa from androgenic anabolic steroids users. J Toxicol EnviroHealth A 2015; 78(2): 67–77.

27- Haveric A, Haveric S, Ibruli S. Micronuclei frequency in peripheral blood and buccal exfoliated cells of young smokers and non- smokers. ToxicolMech Methods 2010; 20 (5): 260-266.

28- Bartsch H, Nair J. Chronic inflammation and oxidative stress in genesis and perpetuation of cancer: role of lipid peroxidation DNA damage, and repair. Lang en becks Arch Sur.2006; 391: 499– 510.

29- Ekbom A, Mc Laughlin JK, Nyren O. Pancreatitis and risk of pancreatic cancer. N. Engl J Med. 1993; 329: 1502–1503.

30- Mondal N, Ghosh S and Ray MR. Micronucleus formation and DNA damage in buccal epithelial cells of indian street boys addicted to gasp "Golden glue". Mutat Res 201; 721;2: 178–183.

31- Visconti R, Grieco D. New insights on oxidative stress in cancer. CurrOpin Drug DiscovDevel. 2009; 12:240–245.

32- Fang J, Seki T, Maeda H. Therapeutic strategies by modulating oxygen stress in cancer and inflammation . Adv Drug Deliv Rev. 2009; 61:290–302.

33- Khandrika L, Kumar B , Koul S, Maroni P, Koul HK. Oxidative stress in prostate cancer. Cancer Lett. 2009; 282: 125–136.

34- Fenech M and Bonassi S. The effect of age, gender, diet, and life style on DNA damage measured using micronucleus frequency in human peripheral blood lymphocytes. Mutagen-esis 2011; 26 (1):43 -49.

35- Gonsebatl ME, Vega AM, Salazar S, et. al. Cytogenetic effects in human exposure to arsenic; Mutat Res 1997; 386 (3): 219–228.

36- Pader M. Oral rinses. Cosmet Toilet Alcohol ClinExp Res. 1994; 109: 59– 6 8.

37- Khan F, Alagappan K, Cardell K. Overlooked sources of ethanol. J Emerg Med 1999;17: 985-988.

38- Rodrigues F, Lehmann M, do Amaral VS, Reguly ML, de Andrade HH. Genotoxicity of threemouthwash products, cepacol, periogard, and plax, in the Drosophila wing-spot test. Environ Mol Mutagen. 2007; 48:644– 649.

39- Talamini R, Vaccarella S, BarbonF, Tavani A, La Vecchia C, Herrero R, Munoz N, Franceschi S. Oral hygiene dentition, sexual habits and risk of oral cancer. Br J Cancer 2000; 83: 1238– 1242.

40- Kristen U, Friedrich RF. Toxicity screening of mouthwashes in the pollen tube growth test: safety assessment of recommended dilutions of twenty brands. In Vivo. 2004; 18:803-807.

41- Winn DM, Blot WJ, Me Laughlin JK, Austin DF, Greenberg RS, Preston-Martin S, Schoenberg JB, Fraumeni JF. Mouthwash use and oral conditions in the risk of oral and pharyngeal cancer . Cancer Res. 1991; 51: 3044- 3047.

42- Allen–Barash N, Wiggins C, Thomans BD, Davis S, Vaughan TL. Effect of mouthwash use on the risk of oral-cancer. Am J Epidemiol. 1993; 138: 635–654 43- Eriksson CJ. Measurement of acetaldehyde: what levels occur naturally and in response to alcohol. Novartis Found Symp.2007;285: 247–255.

44- Bloching M, Hofmann A, Lautenschlager C, Berghaus A, Grummt T. Exfoliated cytology of normal buccal mucosa to predict the relative risk of cancer in the upper aerodigestive tract using the MN-assay.Oral Oncol 2000;36: 550 -555.

45- Kundu JK, Surh YJ. Inflammation: gearing the journey to cancer. Mutat Res. 2008;659:15–30.

### تردد الانوية الصغيرة والاستماتة الخلوية في الخلايا المقشرة لمخاطية الفم لمرضى الالتهاب المزمن لمخاطية الفم

الخلاصة:

الالتهاب المزمن للبطانة المخاطية للفم والذي يحدث بواسطة عوامل بايلوجية او كيميائية او فيزيائية وارتباطه بزيادة مخاطر تحوله الى سرطان الفم.

الهدف من هذه الدراسة تقييم الضرر الوراثي التلقائي الحاصل في خلايا البطانة المخاطية للفم لمرض (Apoptosis) الالتهاب المزمن وذلك بواسطة اختبار النويات الدقيقه (MN) Micronuclei واستماتة الخلايا (Apoptosis) الحاصل في الخلايا المقشرة للبطانة المخاطية للفم.

تتضمن الدراسة الخلوية اضافة الى عد الانوية الصغيرة، عند الاستماتة الخلوية (تمزق الانوية،تكثيف الصبغي،تغلظ) شملت الدراسة (42) فرداً وبعمر يتراوح بين (20 40) سنة منهم (22) فرداً (13 ذكور، 9 اناث) مصابين بالتهاب مزمن للبطانة المخاطية للفم اضافة الى (20) فرداً (12 ذكور، 8 اناث) سليم كمجموعة سيطرة. مصابين بالتهاب مزمن للبطانة المخاطية للفم اضافة الى (20) فرداً (12 ذكور، 8 اناث) سليم كمجموعة سيطرة. حيث كانت زيادة معنوية في تردد (MN) (MN) (20) في خلايا البطانة المخاطية للفم المأخوذة من مناطق الافة حيث كانت زيادة معنوية في تردد (MN) (MN) في خلايا البطانة المخاطية للفم المأخوذة من مناطق الافة (20) وليث كانت زيادة معنوية في تردد (MN) (MN) في خلايا البطانة المخاطية للفم المأخوذة من مناطق الافة (20) المتوسطات (2.4 في المناطق غير المتضررة (Non\_Lesion area) في مرض الالتهاب المزمن للفم حيث كانت المتوسطات (2.40 ± 7.50) وعلى التوالي مقارنة بمجموعة السيطرة حيث كانت (2.40 ± 7.50) والمناطق في تردد الاستماتة الخلوية في مرض الالتهاب المزمن الفم حيث كانت المتوسط (2.40) (2.400) وعلى التوالي مقارنة بمجموعة السيطرة حيث كانت المتوسطات (2.40 ± 7.50) وعلى التوالي مقارنة بمجموعة السيطرة حيث كان المتوسط المتوسطات (2.40 ± 7.50) والمن التوالي مقارنة بمجموعة السيطرة حيث كان المتوسط المتوسطات (2.40 ± 7.50) والمتماتة الخلوية في مناطق الافات المرضى مقارنة بمجموعة السيطرة (2.400)، ولوحظ انخفاض في تردد الاستماتة الخلوية في مناطق الافات المرضى مقارنة بمجموعة السيطرة (2.400).

اما في ما يخص تردد الانوية الصغيرة فيما يتعلق بأرتباطها بمتغيرات (العمر، الجنس، استخدام غسول الفم،صحة الفم).

كان هناك ارتفاع معنوي في تردد (MN) في الاناث مقارنة بالذكور في كلاً من مجموعة المرض والسيطرة (P<0.01) ولم يلاحظ هنالك اي فروق معنوية في وجود (MN) في الفئات العمرية المختلفة.

علاوةً على ذلك كان هنالك زيادة معنوية في تردد (MN) في الاناث اللاتي يعانين من صحة فم غير جيدة واللاتي يستخدمن غسول الفم (P<0.01) اضافة لذلك لوحظ هنالك زيادة في تردد (MN) في عينات المرضى الذين يستخدمون غسول الفم ويعانون من صحة فم غير جيدة،كذلك هنالك ارتفاع معنوي في تردد (MN) عند مستخدمي غسول الفم مقارنة بالذين لايستخدمون غسول الفم في مجموعة السيطرة (P<0.01).

من خلال النتائج نقترح ان زيادة (MN) وتثبط الاستماتة الخلوية ربما ترتبط مع تطور سرطان الفم في مرضى الالتهاب المزمن للبطانة المخاطية للفم.